

SWD Research Update



SWD Non-nutritive sugars project















Can we use non-nutritive sugars to replace toxicants?

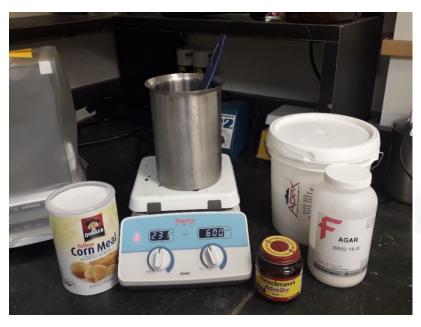




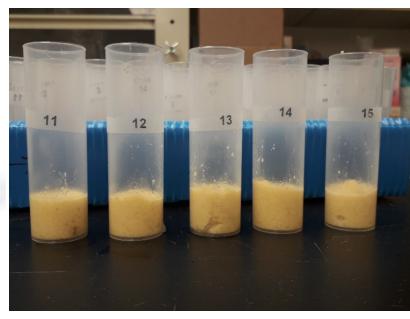
- ➤ Major components of artificial sugars include stevia, erythritol, dextrose, sucralose, saccharin, and aspartame
- ➤ Do non-nutritive sugars have same level of toxicity as insecticide?
- ➤ Can non-nutritive sugars significantly reduce survivorship of SWD?

Replacing Sugar in SWD Diet

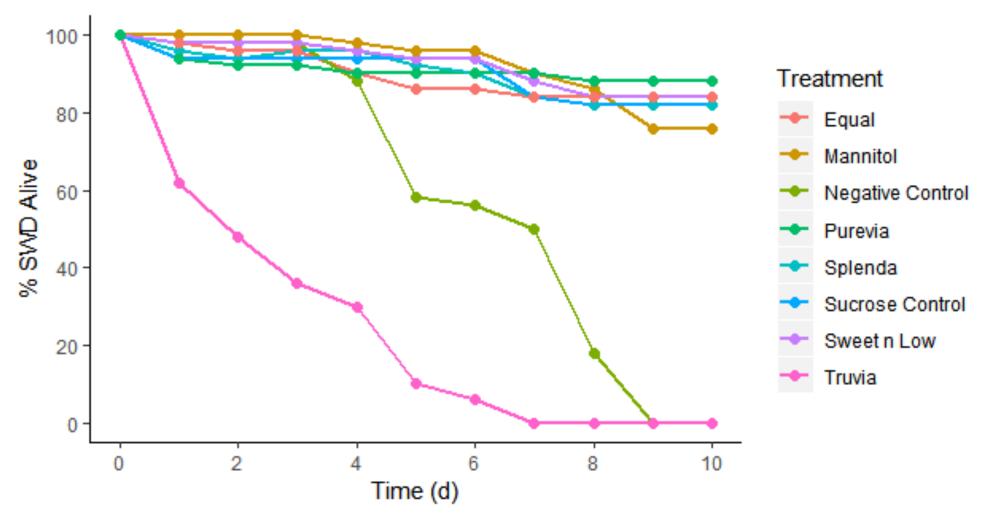
- Prepared Drosophila diet with nutritive sugar component removed
- Replaced sucrose with non-nutritive sugar treatment
- Positive control: sucrose, negative control: water
- Filled Drosophila tubes with 3 cm of diet, and placed 10 adult flies (0-48 hrs old) into each (5 tubes/50 flies per treatment)
- Counted survivorship of each tube daily for 10 days





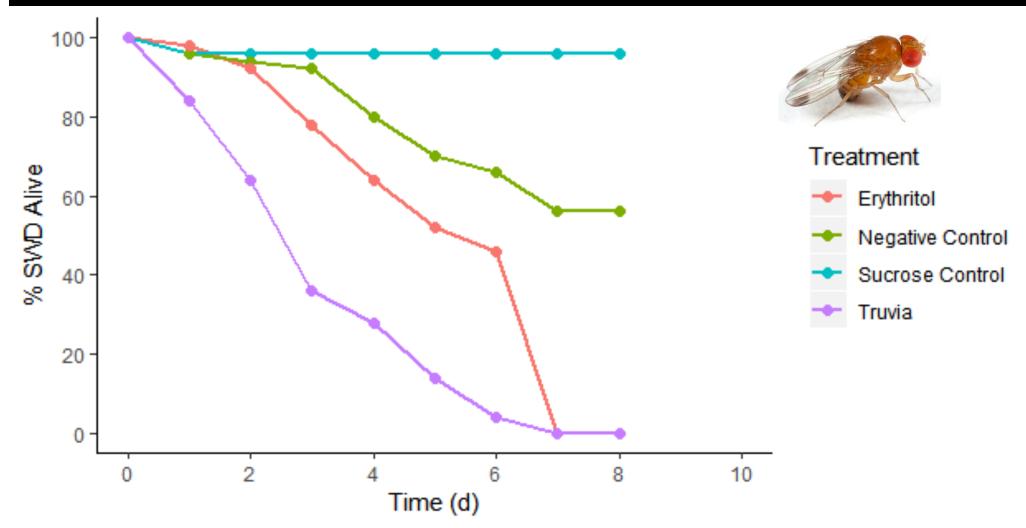


Which commercially available sugars will be toxic to SWD?



Diet with Truvia added in killed SWD significantly faster than all other diets, including the negative control (water) diet

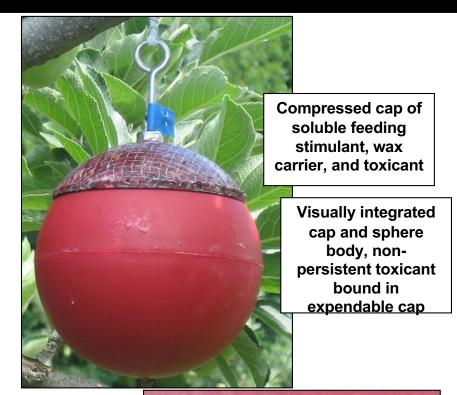
Truvia (erythritol) is toxic to SWD



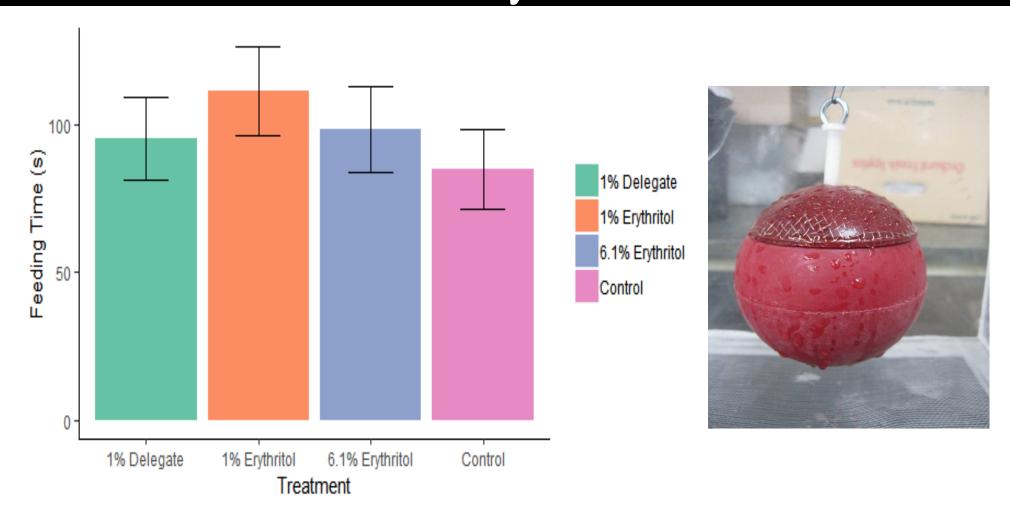
- > Erythritol (a sugar alcohol) is the major component in Truvia
- Kills flies faster than the negative control
- Suggests the erythritol is poisoning rather than starving them

Killing Agent Lethality for SWD

- Evaluate lethality of attracticidal spheres with non-nutritive sugars as toxicant for SWD
- Cap contains a feeding stimulant (sugar) and toxicant
- Exploits environmental moisture (rain and dew) to continuously renew toxicant on sphere surface
- Toxicant not washed away with first rain even or heavy morning dew

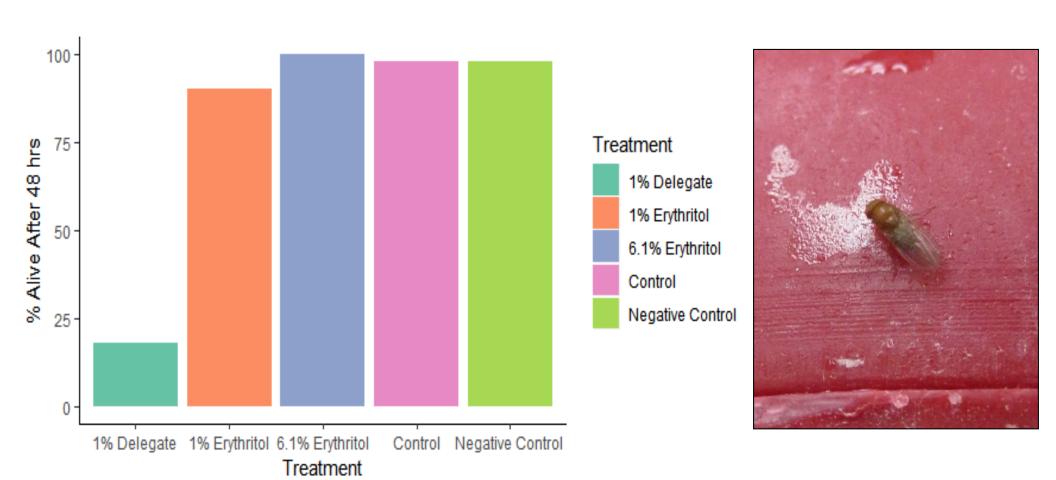


SWD feeding from attracticidal spheres dosed with Erythritol



When SWD were allowed to feed for 5 minutes, there were no significant differences in feeding times among Erythritol, insecticide, and sucrose solution

Survivorship of SWD after feeding from attracticidal sphere for 5 minutes



Only 1% solution of Delegate showed a significant decrease in SWD survival

Tentative Conclusions

- ➤ Erythritol is toxic when included in SWD diet
- ➤ Diet including Erythritol kills SWD faster than sugarfree diets
- ➤ Suggests Erythritol is poisoning rather than starving the flies
- ➤ Erythritol appears to be non-toxic when available for short durations
- ➤ When incorporated into attracticidal spheres, no significant decrease in survival; only delegate showed significant decrease in survival

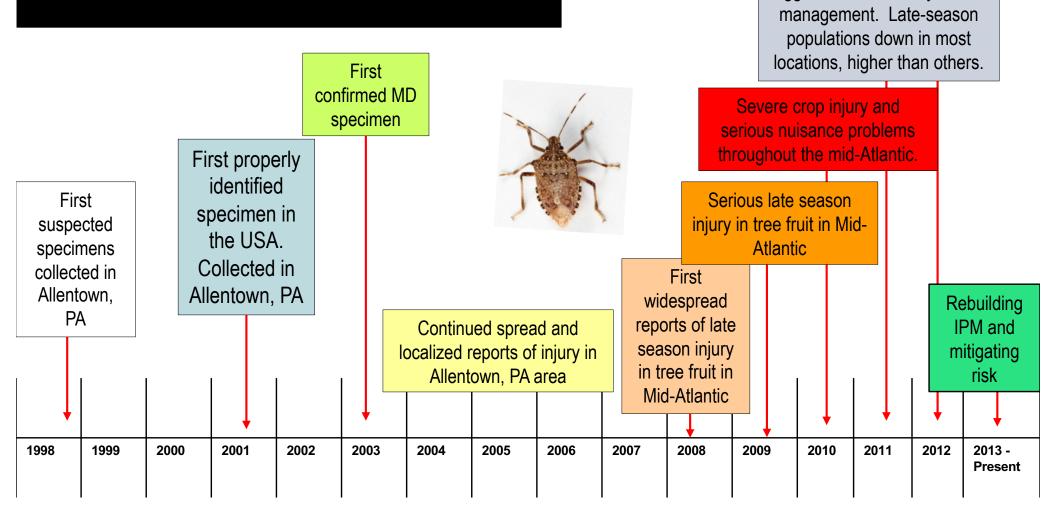




BMSB Research Update

History of BMSB in the

United States

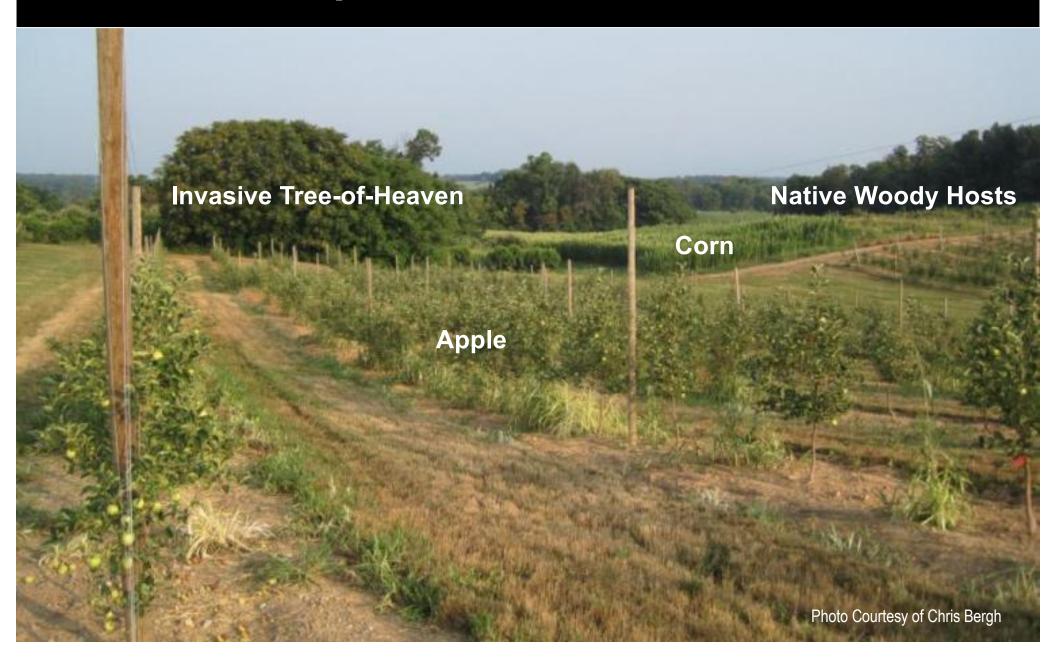


Secondary pest problems become common in east

and increasing populations in west and southeast

Aggressive chemically-based

Landscape-Level Threat To Crops



Can We Develop Reliable Pheromone-Based Monitoring Tools?

- Tools that provide accurate measurements of presence, abundance, and seasonal activity of BMSB
- Inexpensive
- Easy to deploy
- Established thresholds so growers can make informed management decisions and reduce damage levels



Two Approaches To Establishing Thresholds

Retrospective Approach: Establishing Correlations Between Trap Captures and Damage

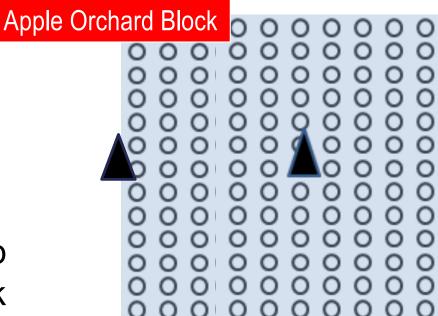
- We found this approach to be problematic
- Many factors that affect captures and damage at harvest
- Non-uniformity among growers (timing and materials) used for spray applications against BMSB and other pests, and delay in injury symptoms appearing leads to a lack of discernable relationship between trap captures and injury

Forward-Driven Approach: Using Set Thresholds To Drive Spray Applications

- This approach establishes that the only sprays applied against BMSB will be triggered by experimental thresholds
- This increases uniformity and enables us to determine if the number of sprays applied at a time indicated by trap captures (based on a set threshold) reduced damage at harvest

Forward-Driven Approach: Establishing A Threshold for Apple

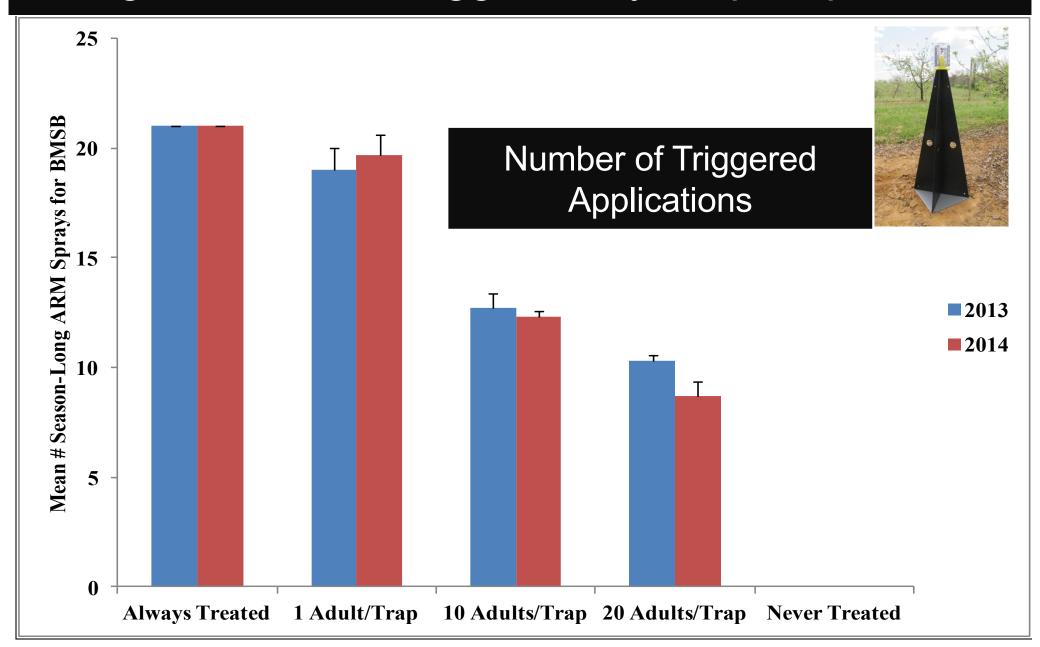
- Apple blocks monitored with two black pyramid traps baited with pheromone lures; traps checked weekly
- When adult captures in either trap reached a set threshold, the block was treated with BMSB material (ARM) and block treated again 7-d later. Threshold was then reset
- This approach enabled the sprays to drive the results against BMSB



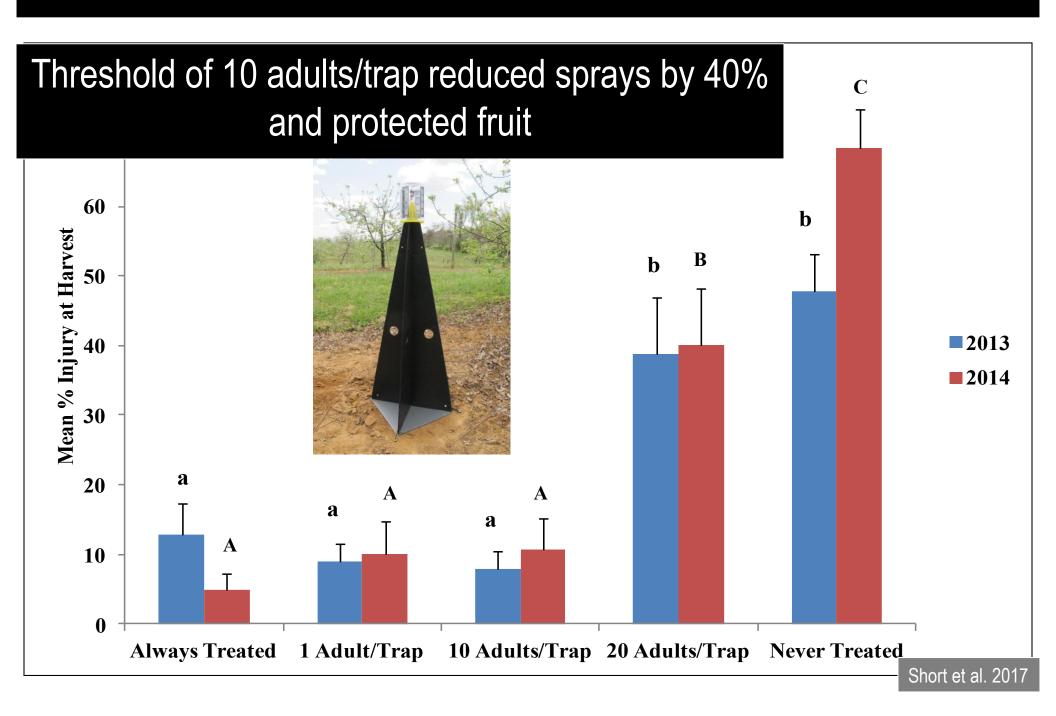
Experimental Treatments

- 1) 1 Adult / Trap
- 2) 10 Adults / Trap
- 3) 20 Adults / Trap
- 4) Treated Every 7 d
- 5) No Spray (Control)

Season-Long Insecticide Applications Made Against BMSB Triggered By Trap Captures



Need for and Timing of Applications Against BMSB



Can We Improve our Trapping System?

- What is the most sensitive and cost-effective trap design and lure formulation?
- Easy to deploy and use?
- Can we detect low populations?
- Can we detect nymphal presence with simplified designs?
- What is the size of the area sampled by the trap?

Targeted Study of Two Trap Designs



Similarities

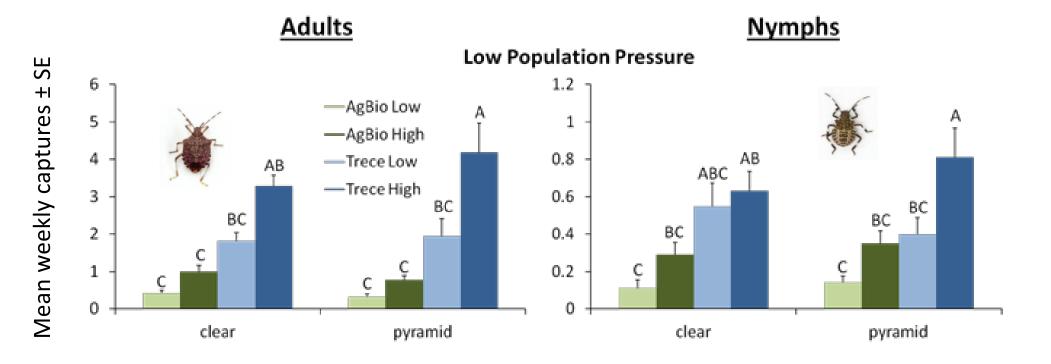
- Ground Deployed
- Upright Visual Stimulus

Differences

- Capture Mechanism
- Retention Mechanism/ Killing Agent



- Trece and AgBio Lures
 - Low: Monitoring dose (1x) (5mg PHER/50 mg MDT)
 - High: Surveillance dose (4x) (20 mg PHER/200 mg MDT)
- Season-long captures of adults and nymphs at 12 sites in the mid-Atlantic



- Trece lure outperformed AgBio lure
- Captures with clear sticky traps statistically similar to pyramid traps
- All traps detected low density BMSB populations
- Nymphs detected with both trap designs

Sensitive Trap-Based Monitoring System



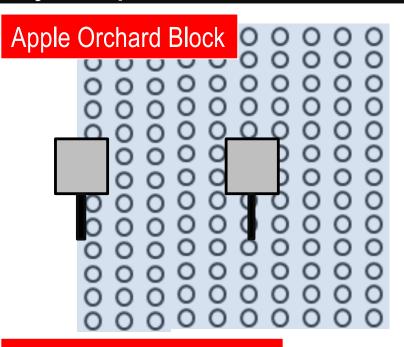
 Capture adults and nymphs at low, moderate or high population levels

 Trap is less expensive and easier to deploy than Black Pyramid Traps

 Trece monitoring lures are longlasting (12 weeks) and sensitive

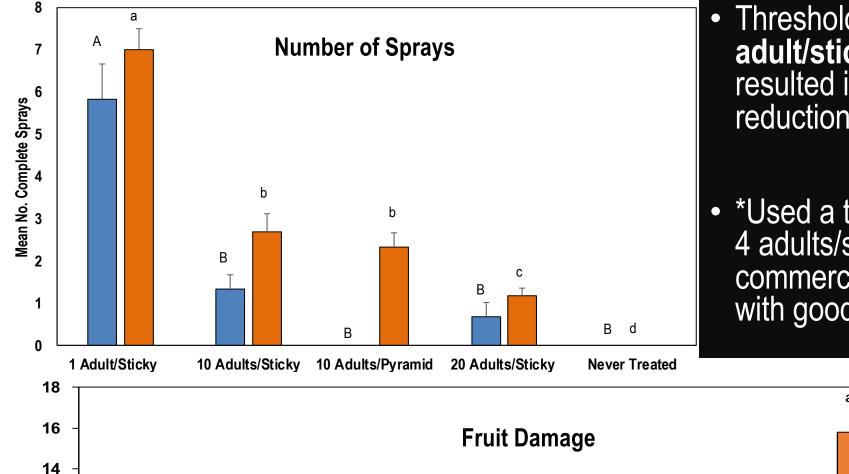
Forward-Driven Approach: Establishing A Threshold for Apple with Clear Sticky Traps

- Apple blocks monitored with two clear sticky panels baited with Trece Dual Lures
- Black pyramid trap standard included
- Traps checked weekly
- When adult captures in either trap reached a set threshold, the block was treated with BMSB material (ARM). Block treated again 7-d later and threshold reset
- This approach enabled the sprays to drive the results against BMSB

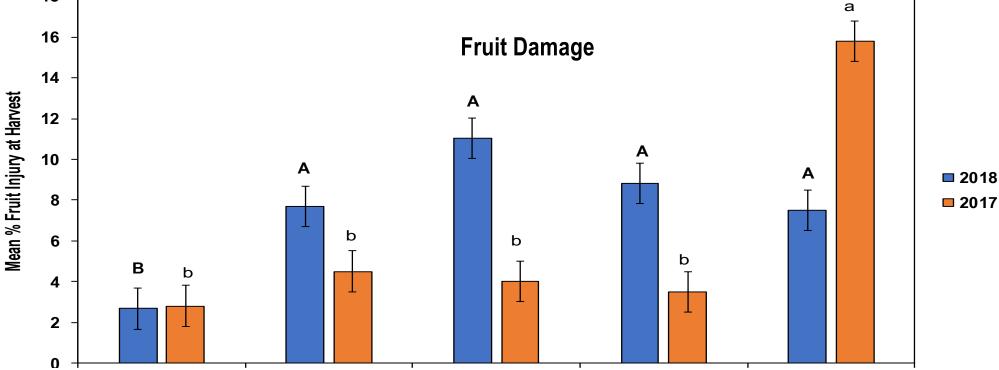


Experimental Treatments

- 1) 1 Adult / Trap
- 2) 10 Adults / Trap
- 3) 20 Adults / Trap
- 4) Treated Every 7 d
- 5) No Spray (Control)



- Threshold of 1
 adult/sticky trap
 resulted in significant
 reductions in injury
- *Used a threshold of 4 adults/sticky trap in commercial orchards with good success

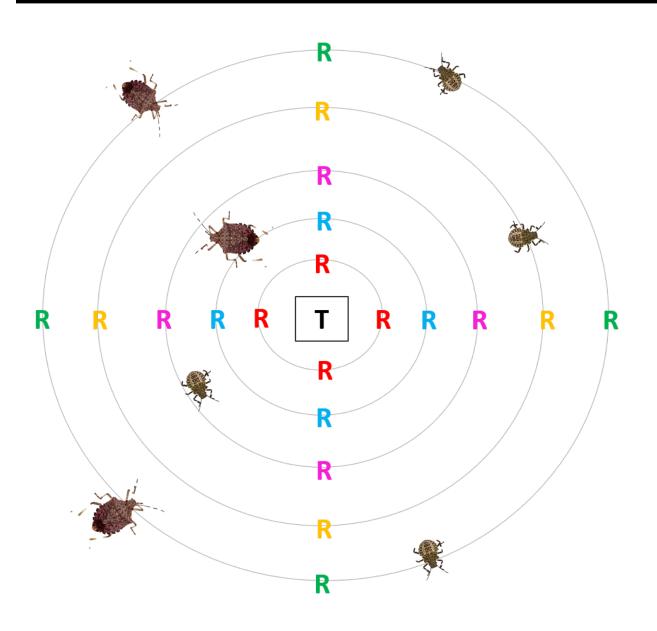


2019 Plans: Establishing A Threshold for Apple Using Clear Sticky Traps

- More work needed to establish accurate threshold
- The following threshold treatments will be evaluated in apple orchards using clear sticky traps baited with Trece Dual Lures
 - 1 adults/sticky trap
 - 4 adults/sticky trap
 - 10 adults/sticky trap
 - Always sprayed (positive control)
 - Never sprayed (negative control)



What is the dispersal capacity of BMSB adults and nymphs?



Single trap, multiple release method



Trapping Theory

50

45

40

35

30

25

20

15

10

5

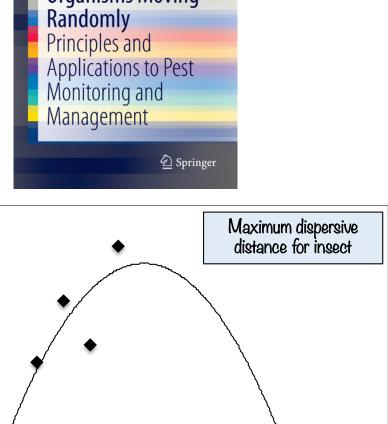
0

0

Proportion Caught*Annulus Area

Christopher G. Adams Jeffrey H. Schenker **Trapping of Small Organisms Moving** Randomly Principles and Applications to Pest Monitoring and Management

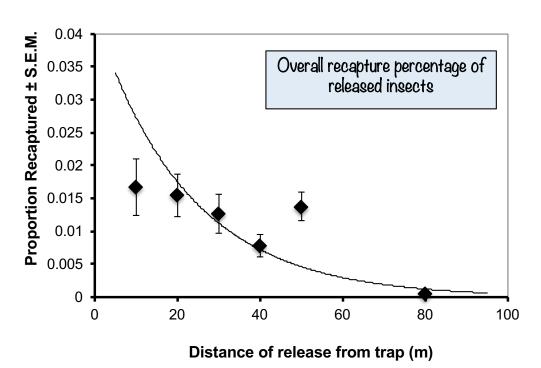
50

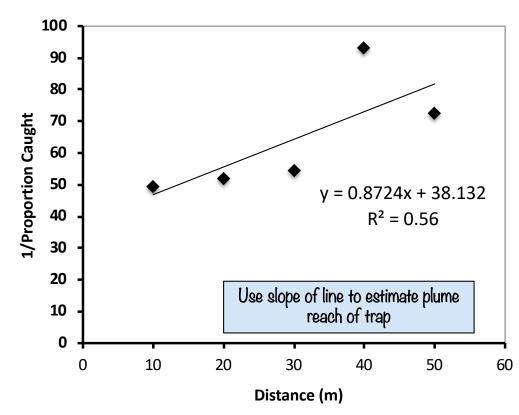


100

Distance (m)

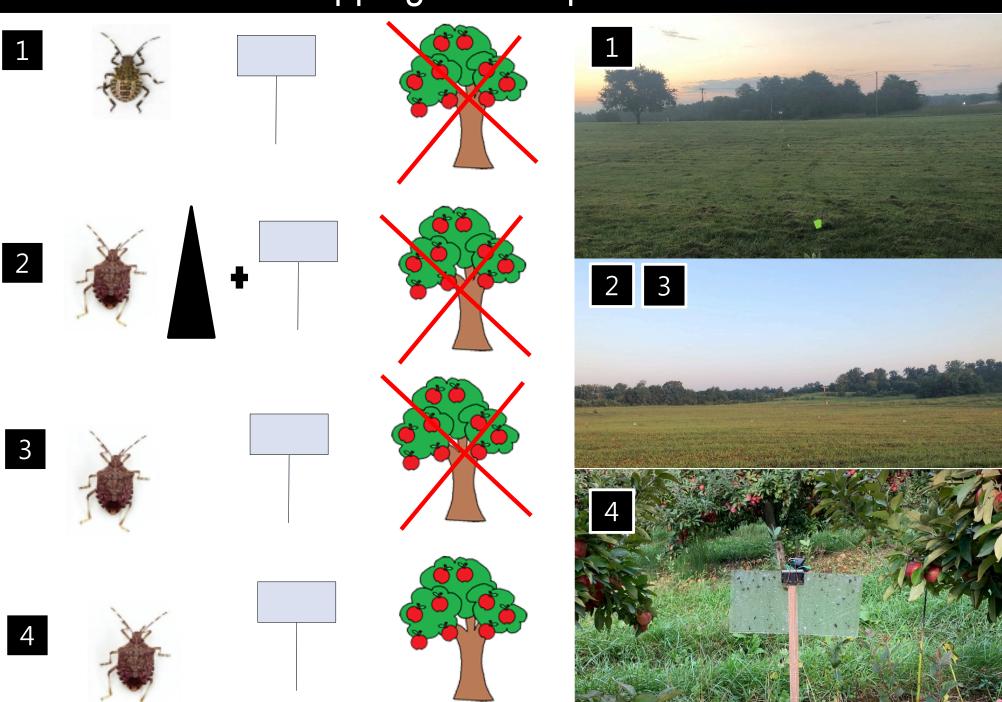
150



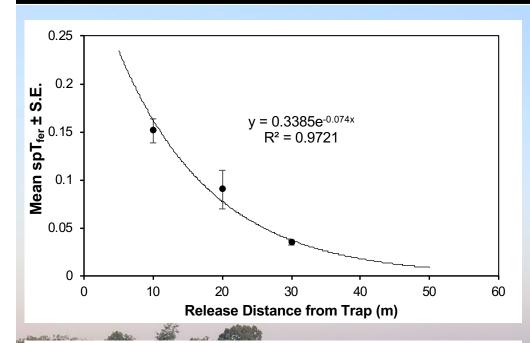


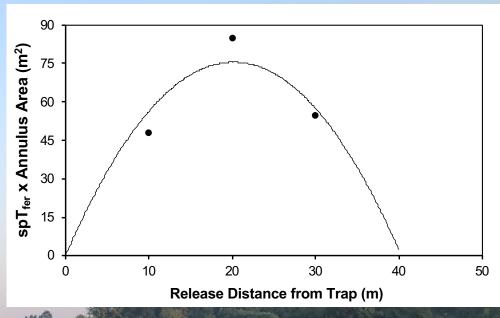


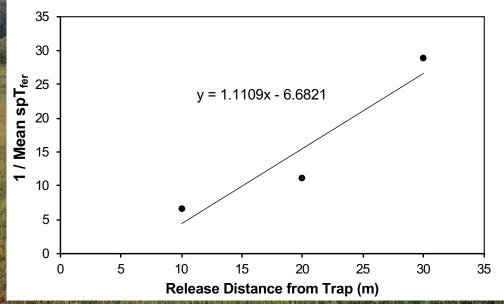
Trapping Area Experiments



Nymphal Trapping Area Results



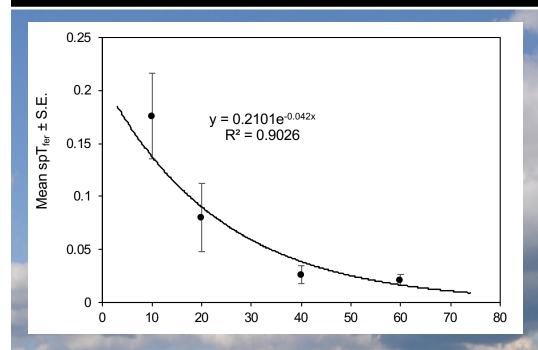


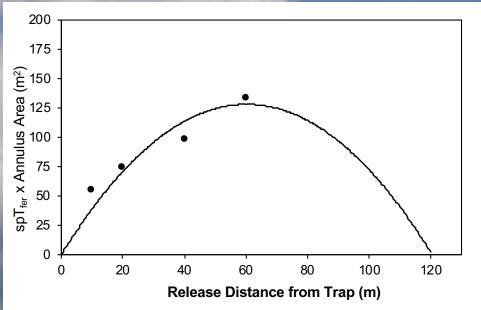


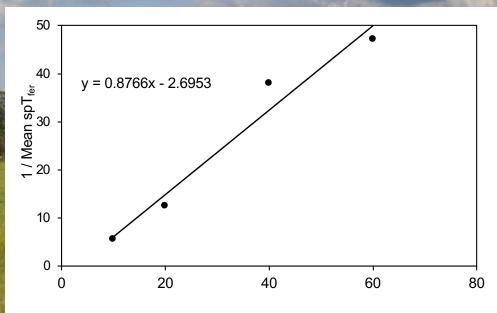
Nymphal Mark-Recapture Results

- Recapture 6.6% of released nymphs
- Maximum Dispersive Distance ~40 m
- Plume Reach < 3 m
- Trapping Radius = 43 m
- Trapping Area = 0.58 ha

Adult Open Field (with Pyramids) Results



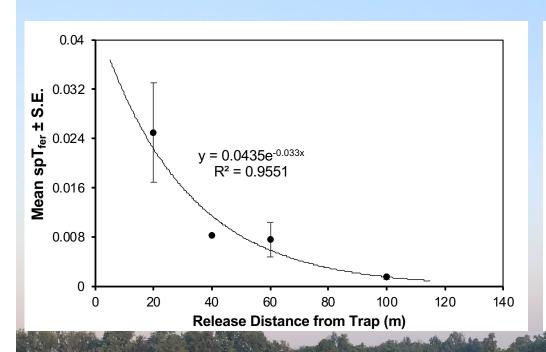


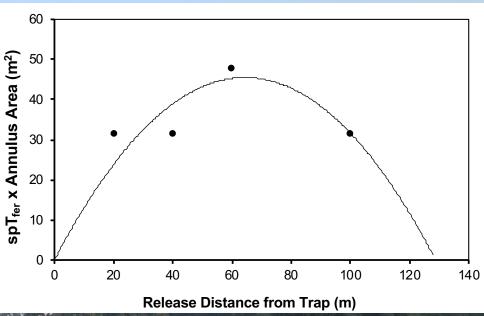


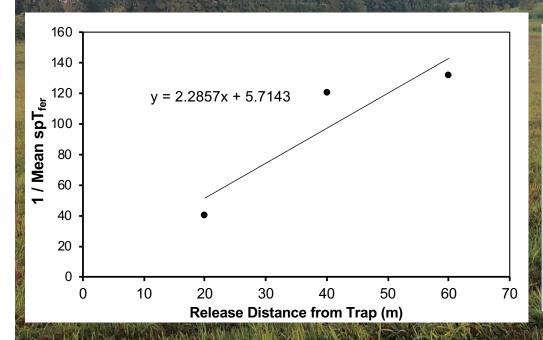
Preliminary Adult Mark-Recapture Open Field

- Overall Recapture Rate: 3.23%
- Max Dispersive Distance ~120 m
- Plume Reach < 3 m
- Trapping Radius = 123 m
- Trapping Area = 4.83 ha

Adult Open Field Trapping Area Results



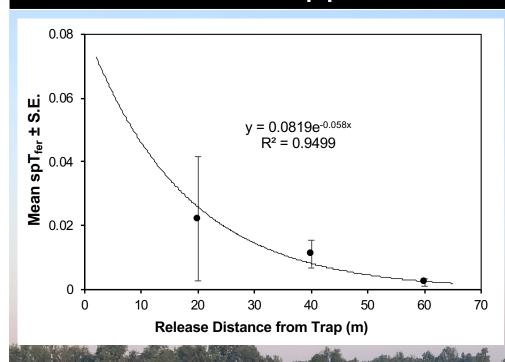


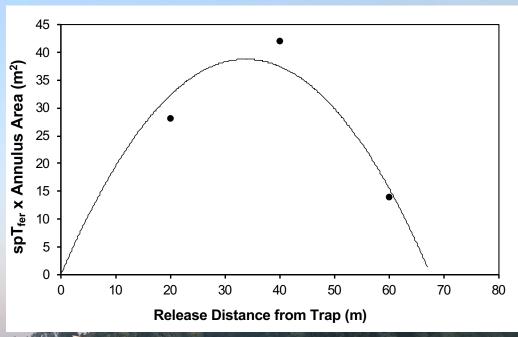


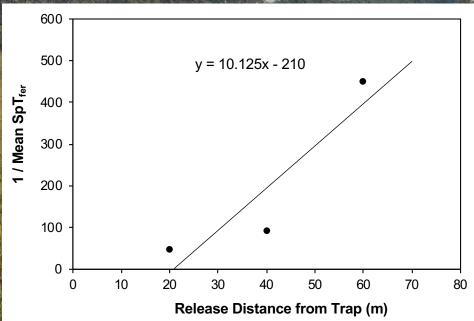
Adults Mark-Recapture Open Field

- Recapture 0.6% of released BMSB
- Max Dispersive Distance ~130 m
- Plume Reach < 3 m
- Trapping Radius = 133 m
- Trapping Area = 5.56 ha

Adult Apple Block Trapping Area Results

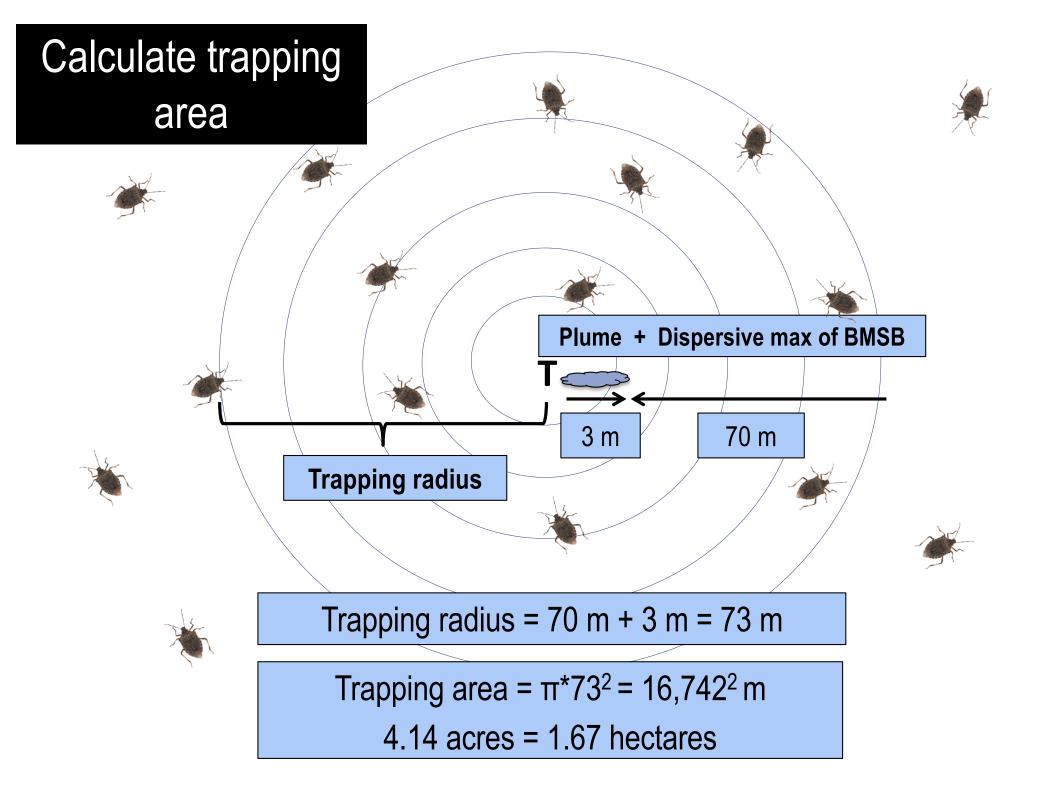






Adults Mark-Recapture Apple Block Edge

- Recapture 1.1% of released BMSB
- Maximum Dispersive Distance ~70 m
- Plume Reach < 3 m
- Trapping Radius = 73 m
- Trapping Area = 1.67 ha



Results For Sticky Panel Trap Baited with Trécé Monitoring Lure

Life Stage	Experiment	Percent Recaptured	Plume Reach	Maximum Dispersal Distance	Trapping Area
Adults	Open Field With Pyramid Traps	3.2%	< 3 m	120 m	4.83 ha
	Open Field	0.6%	< 3 m	130 m	5.56 ha
3 -2	Apple Orchard	1.1%	< 3 m	65 m	1.67 ha
Nymphs	Open Field	6.6%	< 3 m	40 m	0.58 ha

- Adult trapping area in an open field is ~ 5 ha
- •Adult trapping area in an apple orchard is reduced to 1.67 ha
- •Nymphal trapping area is ~0.6 ha; will likely decrease in a host crop
- •Strong behavioral association with host plants that influences response to trap and increases retention time
- •More replication needed in apple orchards and other host crops such as peach, vegetables and field crops to further estimate accurate trapping areas

 Kirkpatrick et al. in prep.

Conclusions and Next Steps

- Non-nutritive sugars for SWD control
 - Erythritol is toxic, but not for short durations of feeding
 - Evaluate erythritol+sugar for attracticidal spheres
- Forward-driven approach to develop management thresholds in apple for BMSB:
 - Pyramid traps: 10 adults/trap protected fruit
 - Clear sticky traps: 1 adult/trap or 10 adult/trap depending on year
 - Future: EVALUATE 1, 4, and 10 adults/trap with always or never sprayed to establish accurate threshold for clear sticky traps
- BMSB adults and nymphs are capable of long range dispersal
 - Estimate of trapping area of ~2 ha for clear sticky traps in apple with one trap placed about every 40 m on apple orchard edge and interior
 - Additional work needed in apples and other crops
- Trapping Area for BMSB changes in the environment in which it is presented
 - Impact of other host plants/vulnerable crops on trapping area?

Acknowledgements



- Demian Nunez
- Layne Leake
- Keirston Rinehart
- Alyssa Lucas
- Brent Short
- Leskey Lab Members

Funding Sources:

- NE SARE # LNE14-334
- Ministry of Primary Industries
- USDA NIFA SCRI # 2016-51181-25409



Thanks!





Danielle M. Kirkpatrick, PhD

Research Entomologist

USDA-ARS Appalachian Fruit Research Station

2217 Wiltshire Road

Kearneysville, WV 25430

Danielle.Kirkpatrick@ars.usda.gov

304-725-3451 x355 (office)

313-418-0540 (cell)



